Instructions for Use

RealLine SARS-Influenza A+B

QUALITATIVE ASSAY KIT FOR THE DIFFERENTIAL DETECTION OF THE SARS-COV-2 RNA WITH TARGETS E-GENE AND N-GENE AND INFLUENZA A AND INFLUENZA B VIRUS RNA BY REAL TIME PCR

RUO – Research use only

The kit consists of two packs, please store **immediately** after delivery:
PART1 at (2 - 8) °C
PART2 at (-18 …-22) °C

RealLine SARS-Influenza A+B (A-Format)	**BI1029-96**	**96 Tests**
RealLine SARS-Influenza A+B (B-Format)	**BI1030-96**	**96 Tests**
valid from:	**October 2020**	

RealLine SARS-Influenza A+B

Explanation of symbols used in labelling

RUO	Research Use Only
LOT	Batch code
REF	Catalogue number
	Contains sufficient for <n> tests
	Use-by-date
	Temperature limit
	Consult instructions for use
	Manufacturer
	Keep away from sunlight

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TABLE OF CONTENT:

Validated Cyclers	
BI1029-96 – A-format	**BI1030-96 – B-format**
RealLine Cycler and equivalent	RealLine Cycler and equivalent Rotorgene Cyler (Qiagen)

RealLine SARS-Influenza A+B

QUALITATIVE ASSAY KIT FOR THE DIFFERENTIAL DETECTION OF THE SARS-COV-2 RNA AND INFLUENZA A AND INFLUENZA B VIRUS RNA BY REAL TIME PCR

1. INTENDED USE

The **RealLine SARS-Influenza A+B Detection Kit** is designed to detect SARS-CoV-2 and the Influenza A and influenza B viruses in human biological samples with the Polymerase Chain Reaction (PCR) method.

Samples are human biological materials: nasopharyngeal swabs, oropharyngeal swabs, bronchoalveolar lavage, endotracheal aspirate, nasopharyngeal aspirate, phlegm.

It is necessary to apply the kit only as directed in this user manual.

Overview to the detected genes and channels:

FAM	HEX	ROX	Cy5
Influenza A-Virus	Internal Control RNA-IC "A"	SARS-CoV-2, E-gene and N-gene	Influenza B-Virus

The extraction of RNA from samples is recommended using the RealLine Prep NA (BI1010, BIORON Diagnostics GmbH).

The **RealLine SARS-Influenza A+B Kit** includes **Internal Control RNA-IC "A",** which is a stabilized RNA molecule. It must be used for quality assessment of the entire assay.

The kit is intended for use with RealLine Cycler (BIORON Diagnostics GmbH), and equivalent cyclers. For the use of Rotorgene Cyclers (Qiagen) the RealLine SARS-Influenza A+B (B-format), REF BI1030-96 with tubes approved should be used.

It is necessary to include the the Negative Control Sample (NC) in every assay. NC is not supplied in this kit but in RealLine Extraction Kits. The Negative control sample is very helpful for contamination control purposes. Use deionized water or sterile buffered saline instead of sample, starting from extraction step.

The use of:

- **Extraction Kits for nucleic acids from clinical specimen from other suppliers**
- **Other real-time PCR devices than described**
- **Appropriate reaction volumes, other than 40 µl**

has to be validated in the lab by the user. Instructions for the use of the Internal Control (IC) have to be followed.

2. KIT CONTENTS

The **RealLine SARS-Influenza A+B** PCR Detection Kit is intended for single use and designed for 96 tests (94 defined samples, one positive control and one negative control).

It is not recommended to perform **less than 8 samples** (6 defined samples, one positive control and one negative control) in one run. It can lead to situation when the volume of enzyme will be insufficient.

Reagent	Quantity	Appearance	Long-time Storage
BI1029-96: RealLine SARS-Influenza A+B (A-format)			
Paraffin-sealed PCR-Mix	12 x 8 strip-tubes, each with 15 µl PCR-mix	Colourless transparent liquid under white wax layer	Box of supplier (2 - 8) °C
SARS-CoV-2 / Influenza RT-PCR-buffer	2 tubes, 810 µl each	Colourless transparent liquid	Box of supplier (2 - 8) °C
Positive Control	1 tube, 130 µl	Colourless transparent liquid	Box of supplier (2 - 8) °C
Internal Control (RNA-IC "A")	1 tube, 1 ml	Colourless transparent liquid	Box of supplier (2 - 8) °C
Strips caps	12 x 8 strip-caps	---	Box of supplier (2 - 8) °C
Enzyme Taq/RT	1 tube, 55 µl	Colourless transparent viscous liquid	in freezer (-18 - -22) °C

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Reagent	Quantity	Appearance	Long-time Storage
BI1030-96: RealLine SARS-Influenza A+B (B-format)			
Paraffin-sealed PCR-Mix	96 tubes, each with 15 µl PCR-mix	Colourless transparent liquid under white wax layer	Box of supplier (2 - 8) °C
RT-PCR-buffer	2 tubes, 810 µl each	Colourless transparent liquid	Box of supplier (2 - 8) °C
Positive Control	1 tube, 130 µl	Colourless transparent liquid	Box of supplier (2 - 8) °C
Internal Control (RNA-IC)	1 tube, 1 ml	Colourless transparent liquid	Box of supplier (2 - 8) °C
Enzyme Taq/RT	1 tube, 55 µl	Colourless transparent viscous liquid	in freezer (-18 - -22) °C

3. PRINCIPLES OF THE PROCEDURE

The method of analysis is based on conducting the reverse transcription of selected fragment of RNA and subsequent amplification of cDNA and detection of PCR products in real time.

Analysis of each specimen is carried out in a separate tube, which contains the Master Mix for RT-PCR.

The reaction is based on a multiplex analysis with a simultaneous detection of multiple targets in one tube.

The Internal Control (IC) RNA is intended to check the quality of extracted RNA used for subsequent reverse transcription in real-time PCR analysis.

The Positive Control plasmid (PC) DNA supplied with the kit is intended for the evaluation of the results of the test.

4. PRODUCT USE LIMITATIONS

- Diagnostic sensitivity of the kit may vary depending on the pathogen prevalence, insufficiency of patient sample and characteristics of the enrolled cohort.
- Reliable results depend on adequate specimen sampling.
- Positive results indicate active or/and asymptomatic infection; results should be interpreted with consideration of clinical and laboratory findings.
- Negative results indicate lack of detectable RNA but do not exclude the infection or disease.
- Potential mutations within the target regions of the SARS-CoV-2 virus and the Influenza A and B virus genome covered by the primers and/or probes used in the kit may result in failure to detect the presence of the pathogens.
- Using of results in combination with COVID-19 lies in the responsibility of the user and clinicians.
- The kit is intended to be used for the detection of SARS-CoV-2 virus RNA, Influenza A virus RNA and Influenza B virus RNA and should be interpreted with consideration of clinical and laboratory findings.

The detection result of this product is only for clinical reference, and it should not be used as the only evidence for clinical diagnosis and treatment. The clinical management of patients should be considered in combination with their symptoms/signs, history, other laboratory tests and treatment responses. The detection results should not be directly used as the evidence for clinical diagnosis, and are only for the reference of clinicians.

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5. SPECIFICATIONS

5.1. Sensitivity:

Analytical sensitivity of the **RealLine SARS-Influenza A+B** Kit is 10 copies of RNA per amplification tube. Sensitivity is determined by the analysis of serial dilutions of the laboratory control sample (LCS).

Sensitivity depends on the sampling and the final volume of the extracted NA (elution volume). Sensitivity of 10 copies per amplification tube corresponds to the following values of the RNA concentration in the sample in case of using RealLine Prep NA (REF BI1010, BIORON Diagnostics GmbH): 1000 copies/ml sample.

5.2. Specificity:

The analytical specificity of the **RealLine SARS-Influenza A+B** Kit was assessed by bioinformatic analyses using available on-line databases with up-to-date comprehensive genetic information. The specific oligonucleotides used in the test were checked against GenBank database sequences. None of the sequences showed sufficient similarity for unspecific detection.

There are no non-specific positive results of amplification of RNA sample in the presence of:

Human coronaviruses HKU-1, NL-63, OC-43, 229E,
Human parainfluenza viruses type 1-4,
Human respiratory syncytial virus,
Human metapneumovirus,
Human rhinovirus,
DNA of Human adenovirus,
Human bocavirus,
Mycoplasma pneumoniae,
Streptococcus pneumoniae,
Chlamydophila pneumoniae,
Haemophilus influenzae,
Klebsiella pneumoniae,
Moraxella catarrhalis,
Bordetella pertussis,
Bordetella parapertussis,

as well as human DNA in concentrations up to 1.0×10^8 copies/ml of the sample.

6. WARNING AND PRECAUTIONS

- Handle and dispose all biological samples, reagents and materials used to carry out the assay as if they were able to transmit infective agents.
- The samples must be exclusively employed for certain type of analysis. Samples must be handled under a laminar flow hood.
- Tubes containing different samples must never be opened at the same time.
- Pipettes used to handle samples must be exclusively employed for this specific purpose. The pipettes must be of the positive dispensation type or be used with aerosol filter tips. The tips employed must be sterile, free from the DNases and RNases, free from DNA and RNA.
- The reagents must be handled under a laminar flow hood.
- The reagents required for amplification must be prepared in such a way that they can be used in a single session.
- Pipettes used to handle reagents must be exclusively employed for this specific purpose. The pipettes must be of the positive dispensation type or be used with aerosol filter tips. The tips employed must be sterile, free from the DNases and RNases, free from DNA and RNA**.** Avoid direct contact with the biological samples reagents and materials used to carry out the assay.
- Use a powder-free surgical gloves.
- Avoid producing spills or aerosol.
- Any material coming in contact with the biological samples must be treated for at least 30 minutes with disinfecting solution or autoclaved for 1 hour at 121 ºC before disposal.
- Molecular biology procedures, such as nucleic acids extraction, reverse transcription, amplification and detection require qualified staff to avoid the risk of erroneous results, especially due to the degradation of nucleic acids contained in the samples or sample contamination by amplification products.
- All oligonucleotide components are produced by artificial synthesis technology according to internal quality control protocol and do not contain blood or products of blood processing.
- Positive control is produced by artificial synthesis technology. Positive control does not include parts of infectious agents.
- All the liquid solutions are designed for single use and cannot be used more than once in amplification reactions.
- Plastic tubes do not contain phthalates.
- Do not breathe gas/fumes/vapor/spray produced by the components of the kit.
- Do not eat/drink components of the kit. Avoid contact with eyes. Only use the reagents provided in the kit and those recommended by manufacturer.
- Do not mix reagents from different batches.
- Do not use reagents from third party manufacturers' kits.
- All laboratory equipment, including dispensers, test tube racks, laboratory glassware, lab coats, bouffant caps, etc., as well as reagents should be strictly stationary. It is not allowed to move them from one room to another. Equip separate areas for the extraction/preparation of amplification reactions and for the amplification/detection of amplification products.

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- Never introduce an amplification product in the area designed for extraction/preparation of amplification reactions.
- Wear lab coats, gloves and tools, which are exclusively employed for the extraction/preparation of the amplification reaction and for the amplification/detection of the amplification products.
- Never transfer lab coats, gloves and tools from the area designed for amplification/detection of the amplification products to the area designed for extraction/preparation of amplification reactions.
- Amplification products must be handled in such a way as to reduce dispersion into the environment as much as possible, in order to avoid the possibility of contamination.
- Pipettes used to handle amplification products must be exclusively employed for this specific purpose.
- Remove PCR waste only in a closed form. Do not open the tubes after amplification.
- Waste materials are disposed of in accordance with local and national standards. All surfaces in the laboratory (work tables, test tube racks, equipment, etc.) must be treated daily with disinfecting solution.

Emergency actions

Inhalation:

Inhalation of the Master Mix contained within this kit is unlikely, however care should be taken.

Eye Contact: If any component of this kit enters the eyes, wash eyes gently under portable running water for 15 minutes or longer, making sure that the eyelids are held open. If pain or irritation occurs, obtain medical attention.

Skin Contact: If any component of this kit contacts the skin and causes discomfort, remove any contaminated clothing. Wash affected area with plenty of soap and water. If pain or irritation occurs, obtain medical attention.

Ingestion: If any component of this kit is ingested, wash mouth out with water. If irritation or discomfort occurs, obtain medical attention.

- Do not use the kit:

If the transportation and storage conditions are breached;

If the reagents' appearance does not respond to the kit passport;

If the kit components packaging is breach;

After the expiry date provided.

- **Significant health effects are NOT anticipated from routine use of this kit when adhering to the instructions listed in the current manual.**

7. EQUIPMENT AND MATERIALS REQUIRED BUT NOT SUPPLIED

Specimen collection

Specimen collection swabs: use only dacron, rayon, or calcium alginate tipped collection swabs with plastic or non-aluminium wire shafts;

RNA extraction and PCR

Specimen and control preparation

- Biological safety cabinet class II;
- Vortex mixer;
- Refrigerator;
- Nucleic acid extraction kit (RealLine Prep NA Kit is recommended);
- High speed centrifuge (RCF 13000 x g);
- Solid-state thermostat (temperature range 40-95°C);
- PCR tube rack for 1.5 ml tubes;
- 1.5 ml microcentrifuge tubes with caps;
- Physiological saline solution 0.9 % NaCl (Sterile);
- Container for used pipette tips;
- Electric laboratory aspirator with trap flask for the removal of supernatant;
- RNase and DNase free filtered pipette tips for aspirator with trap flask;
- Powder-free surgical gloves;
- Disinfectant solution.

Pre-amplification-reagent preparation area

- UV PCR cabinet;
- Vortex mixer;
- Refrigerator;
- PCR tube rack for 0.2 ml tubes or strips;
- Rotor for strips (if package in strips is used);
- Single channel pipettes (volume range 2-20 µl, 20-200 µl, 200-1000 µl);
- RNase and DNase free filtered pipette tips (volume range 20 µl, 200 µl, 1000 µl);
- Powder-free surgical gloves;
- Disinfectant solution;

Post-Amplification – Amplification detection area

- Real-Time PCR Cycler

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8. SAMPLES

The **RealLine SARS-Influenza A+B Kit** is designed to detect RNA extracted from the nasopharynx and oropharynx swabs, bronchoalveolar lavage, endotracheal aspirate, nasopharyngeal aspirate, sputum, depending on professional prescription.

Interfering substances

The presence of PCR inhibitors in a sample may cause controversial (uncertain) results. The sign of PCR inhibition is the simultaneous absence of signal in internal control and specific product of amplification.

PCR inhibitors are the presence of haemoglobin in the RNA sample as a result of incomplete removal during the extraction of RNA from a biomaterial sample containing an impurity of blood, as well as the presence of isopropyl alcohol and methyl acetate in the extracted RNA sample as a result of incomplete removal of washing solutions during sample preparation.

The maximum concentration of interfering substances, which do not affect the amplification of the laboratory control sample and internal control: haemoglobin – 0.35 mg/ml cDNA sample, isopropyl alcohol – 100 µl /ml cDNA sample, methyl acetate – 100 µl/ml cDNA sample.

To reduce the potential PCR inhibitors, it is necessary to follow the principles of taking biological material. If suspecting a large count of PCR inhibitors in the sample, it is recommended to choose NA extraction methods that allow to remove PCR inhibitors from the sample as much as possible. It is not recommended to use express methods of NA extraction.

The features of biomaterial sampling

Work with biomaterials should be performed in accordance with Laboratory testing for coronavirus disease (COVID-19) in suspected human cases, Interim guidance, 19 March 2020 (WHO) and national legislation.

The collection of clinical material and its packaging is carried out by an employee of a medical organization who is trained in the requirements and rules of biological safety when working and collecting material suspected of being infected with pathogenic microorganisms.

The timing of biomaterial sampling is very important. Presumably, the highest content of the virus in the respiratory organs of person can be within the first 4 days after the appearance of symptoms of the disease. Samples should be collected within 3 days after the appearance of clinical symptoms of the disease.

At least three types of clinical material should be collected from one patient.

It is necessary to take swabs from the nasal cavity, naso-and oropharynx.

Each sample of biomaterial should be placed in a separate transport container.

Transportation and storage of the samples

Type of the sample	Collecting material requirements	Transportation	Storage conditions before transportation	Comments
Nasopharynx and oropharynx swabs	Plastic test tubes and tampons for swabs	4 °C	≤5 days: 4 °C >5 days *: -70 °C	Nasopharyngeal and oropharyngeal tampons should be placed in the same tube to increase the viral load
Bronchoalveolar lavage	Sterile container	4 °C	≤48 hours: 4 °C >48 hours *: -70 °C	A small sample dilution is possible
Endotracheal aspirate, nasopharyngeal aspirate or nasal lavage	Sterile container	4 °C	≤48 hours: 4 °C >48 hours *: -70 °C	
Sputum	Sterile container	4 °C	≤48 hours: 4 °C >48 hours *: -70 °C	Make sure that the material is from the lower respiratory tract

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9. PROCEDURE

9.1. RNA-Extraction

 *Regardless of the nucleic acid extraction kit used, a negative control sample must underwent all RNA extraction procedure stages (a sterile physiological solution or transport medium for clinical samples in the volume specified on the user manuals for the nucleic acid extraction kit can be used).*

The Internal control **RNA-IC "A"** from the **RealLine SARS-Influenza A+B** kit must be used as an Internal Control sample.

- Mark 1.5 ml tubes according to the number of specimen and NC
- Add 10 µl of **RNA-IC "A"** to each sample including NC

Perform the RNA Extraction procedure according the user manual supplied with the respective kit.

⚠ *The resulting RNA preparation must be used immediately for RT-PCR. If it is needed the resulting RNA preparation can be stored at temperature from (-18…-22) °C for no longer than a week with a single defrost before reverse transcription. But be aware: the direct use of freshly prepared RNA will bring a more reliable result, especially with low RNA contents*

9.2. PCR Amplification with Reverse Transcription (RT-PCR)

 *The reagents and tubes should be kept away from direct sun light.*

1. Mark the required number of tubes with paraffin sealed RT-PCR-mix according to the number of samples to be analysed, 1 tube for Positive Control **PC** and 1 tube for Negative Control **NC**

Example: if you need to test 2 samples, mark 2 tubes for samples, 1 tube for **PC** and 1 tube for **NC**. Total number of tubes – 4.

2. Vortex the **RT-PCR-buffer** and **Enzyme Taq/RT** thoroughly for 3-5 s, then spin briefly for 1-3 s.

⚠ *Enzyme Taq/RT should be take out from the freezer immediately prior to use.*

3. Prepare the mixture of SARS-CoV-2/Influenza **RT-PCR-buffer** and **Enzyme Taq/RT**. Add to one tube:

15 x (N+1) **µl** of SARS-CoV-2/Influenza **RT-PCR-buffer**,

0.5 x (N+1) **µl** of **Enzyme Taq/RT**,

Where: N – is the amount of the samples, including PC and NC.

4. Vortex the tube, then spin briefly for 1-3 s.

⚠ *Mixture of RT-PCR-buffer and Enzyme Taq/RT should be prepared immediately before use, it must be used within 2 hours after preparation, storage at temperatures from (2 – 8) °C.*

5. Add **15 µl** of Enzyme Taq/RT and -RT-PCR-buffer **mix** into the each PCR tube. Avoid paraffin layer break.

6. Vortex the tubes with samples and PC and NC for 3-5 s and spin down the drops by centrifuging for 1-3 s.

⚠ *Please be carefully, prevent contamination and use filter tips.*

7. Add **10 µl** of **RNA sample** into corresponding RT-PCR tube. Close the tubes tightly. Add **10 µl** of **NC** which passed the whole NA extraction procedures into corresponding tube. Add **10 µl** of **PC** into **PC** tube. Close the tubes tightly. Avoid paraffin layer break.

Reagent	Patient Sample(s)	PC	NC
Extracted RNA	10 µl	-	-
NC	-	-	10 µl
PC	-	10 µl	-

8. Spin the tubes briefly for 3-5 s to collect the drops.

9. Place the tubes into the Thermal Cycler

For the RealLine Cycler (BIORON Diagnostics GmbH), we can provide the PCR program for easy use on request: techsupport@bioron.de

Launch the RealTime_PCR application in “Device operation” mode. Upload the **“.ini”** file before the first run. Add test in subsequent runs. Specify the number and identificator of samples. Define position of tubes in software interface according to position they were set in thermal unit. Run PCR.

If you use another Real-Time PCR cycler, please contact us for support: techsupport@bioron.de

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10. Program the RealLine Cycler as follows

Step 1:	35 °C	20 min	1 cycle
Step 2:	95 °C	5 min	1 cycle
Step 3:	94 °C	10 s	50 cycles*
	64 °C*	15 s	
Step 4	80 °C*	1 s	1 cycle
Step 5	10 °C*	Hold	
* measurement of fluorescence			

For Rotorgene Cyclers

Step 1:	32 °C	20 min	1 cycle
Step 2:	95 °C	5 min	1 cycle
Step 3:	94 °C	10 s	50 cycles*
	60 °C*	15 s *	
* measurement of fluorescence			

11. Choose channels:

FAM / ***Green***	**HEX** / ***Yellow***	**ROX** / ***Orange***	**Cy5** / ***Red***
Influenza A-Virus	Internal Control RNA-IC “A”	SARS-CoV-2, E-gene and N-gene	Influenza B-Virus

12. PCR Volume: **40 µl**

13. Match the positions of test tubes with samples, positive and negative controls according to the instruction manual for the real time PCR system in use.

14. Run the program.

10. CONTROLS

The **RealLine SARS-Influenza A+B Kit** contains **Positive Control** sample **PC**. Positive control is a cloned part of the virus genome. It is produced with genetic engineering techniques and characterized by automatic sequencing.

The kit includes **the Internal control (RNA-IC).** RNA-IC is intended to assess the quality of the RNA extraction and polymerase chain reaction. To reveal possible contamination a negative control is required.

A **Negative Control NC** sample should go through all stages of RNA extraction. Physiological saline solution can be used as a negative control sample in volumes indicated in supplied instructions.

The test result is considered valid if:

- the exponential growth of the fluorescence level for the specific product is present, in this case the internal control is not taken into account;
- the exponential growth of the fluorescence level for the specific product is absence and for internal control is present.

The test result is considered **invalid** when the exponential growth of the fluorescence level for the specific product and for internal control are not observed.

If **Positive Control (PC)** does **not** express growing fluorescence of the specific product or positive result, it is required to repeat the whole test. It may be caused by inhibitors, operation error or violation of storage and handling.

If **Negative control (NC)** expresses growing fluorescence of the specific product or positive result, all tests of the current batch are considered false. Decontamination is required.

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11. DATA ANALYSIS

In case of using Real-Line Cyclers, the analysis is performed automatically. In all other cases, the analysis is based on the presence or absence of specific signal.

The Real-time PCR Thermal Cyclers detects and interprets results automatically. Analysis will be performed by Real-Time PCR application.

PCR results interpretation should be carried out in accordance with following Table

Detection Channel				Interpretation
FAM/*Green*	**HEX/*Yellow***	**ROX/*Orange***	**Cy5/*Red***	
Influenza A	**RNA IC**	**SARS-CoV-2 E-gene/N-gene**	**Inluenza B**	
Analyzed samples				
+	Not considered	**-**	**-**	**RNA of Influenza A is detected**
-	Not considered	**+**	**-**	**RNA of SARS-CoV-2 is detected**
-	Not considered	**-**	**+**	**RNA of Influenza B is detected**
-	**+**	**-**	**-**	RNA of SARS-CoV-2 virus, Influenza A and Influenza B virus is not detected
-	**-**	**-**	**-**	Unreliable result! Repeat NA extraction and PCR amplification or re-collect of a clinical sample performed sequentially
Positive Control sample				
+	**-**	**+**	**+**	**Positive result**
Negative Control sample				
-	**+**	**-**	**-**	**Negative result**

“ + “: Cp is specified **“ – “ Cp is not specified**

Unreliable results may be caused by the presence of inhibitors in the nucleic acid preparation obtained from the clinical material, errors in the pre-analytical stage, incorrect implementation of the analysis Protocol, non-compliance with the temperature mode of amplification, etc. In this case, either re-staging of reverse transcription and polymerase chain reaction, or re-extracting of the nucleic acid preparation, or re-collect of clinical material (performed sequentially) is required.

When growing fluorescence (Cp/Ct is specified) through the Fam/Green, Rox/Orange, or Cy5/Red channels is expressed for Negative Control (NC), the results of whole series are considered false. It is required to eliminate contamination.

The controls should be also considered to exclude false positive and false negative results (see Paragraph 10 of the current manual).

Please use the **cut-off Ct ≤ 40** (specified Product) and **33 (PC)** for Rotor-Gene thermal cycler. These cut-offs can be recommended for other cyclers, but depends on your complete analytical setup.

The result characterized by Ct above this value should be considered doubtful and the whole assay should be repeated.

12. STORAGE AND TRANSPORTATION

- Expiry date – 12 months from the date of production.
- Both parts of the kits must not delivered and keep with dry ice
- All components of **RealLine SARS-Influenza A+B Kit PART 1** must be stored out of light at temperatures from (2 – 8) °C during the storage period. Excessive temperatures and light can be detrimental to product performance.
- The Enzyme Taq/RT, component in **RealLine SARS-Influenza A+B Kit PART 2** must be stored at temperatures from (- 18°C to – 22) °C during the storage period.
- The kit can be transported by all types of roofed transport at temperatures corresponding to the storage conditions of the kit components over the transportation. Transportation is allowed in thermal containers with icepacks by all types of covered transport at a temperatures up to 25 °C inside the container, but for no longer than 5 days.
- An expired **RealLine SARS-Influenza A+B Kit** should not be used
- It is strongly recommend to follow the given instructions in order to obtain accurate and reliable results.
- Do not use kits with damaged inner and outer packaging and get in contact with BIORON Diagnostics GmbH.

Shelf-life of the kit following the first opening of the primary container:

- Components of the kit should be stored at temperatures from (2 – 8) °C during the storage period;
- PCR-mix for amplification should be stored at temperatures from (2 – 8) °C and out of light during the storage period; optimally in the foil pouches;
- Enzyme Taq/RT should be stored at temperatures from (- 18°C to – 22) °C during the storage period.

Please avoid freeze/thaw cycles of the Enzyme Taq/RT component.

The conformity of the **RealLine SARS-Influenza A+B Kit** to the prescribed technical requirements is subject to compliance of storage, transportation and handling conditions recommended by manufacturer.

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13. TROUBLESHOOTING

	Possible cause	Solution	Result
PC	-	Operation error PCR inhibition Violation of storage and handling requirements	Repeat whole test Dispose current batch
NC	+	Contamination	Dispose current batch Perform decontamination procedures
IC	-	PCR inhibition RNA extraction violation	Repeat RNA extraction Repeat whole test Resample

If you face any undescribed issue, Remarks, requests and comment, please contact: techsupport@bioron.de